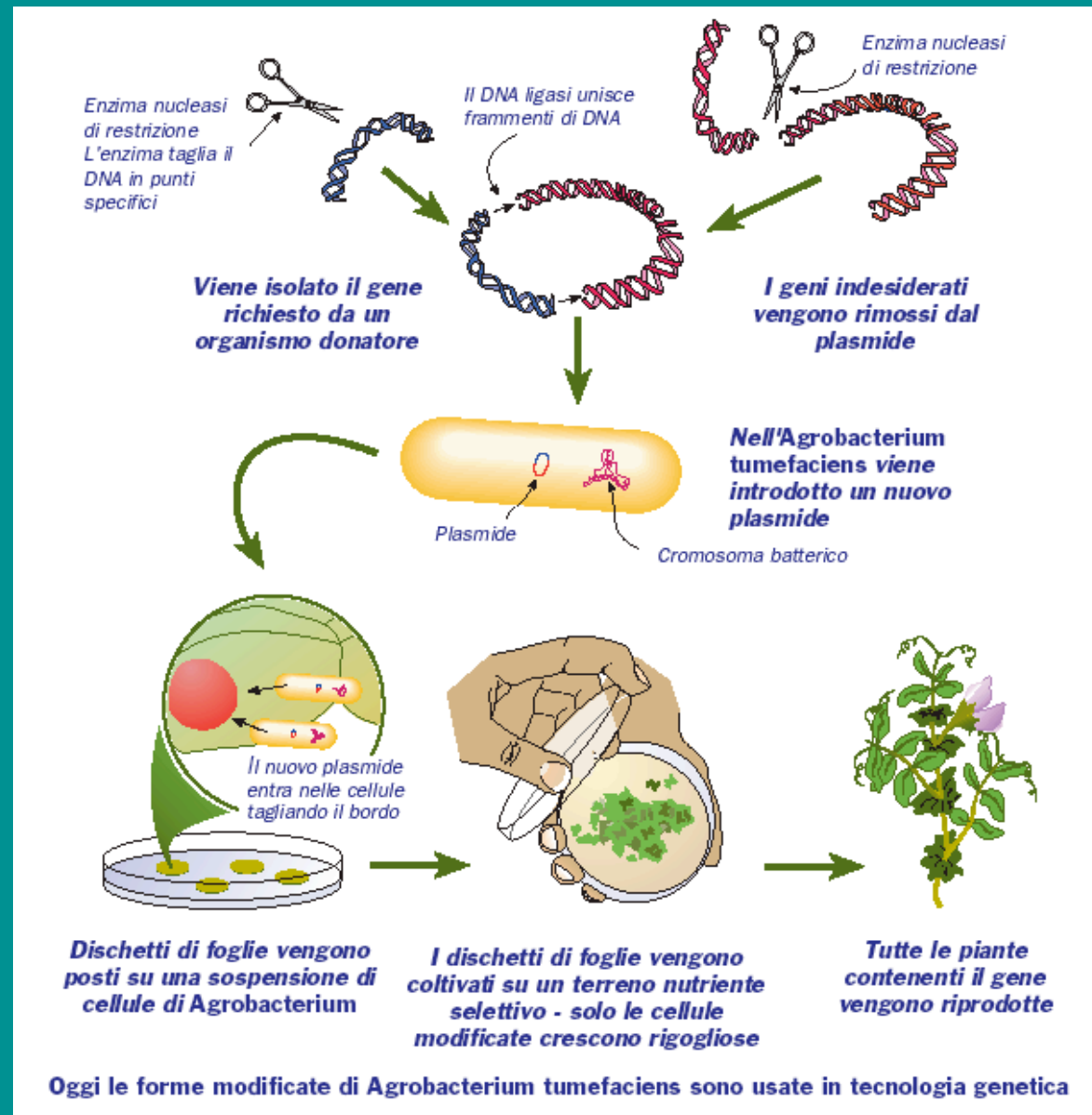
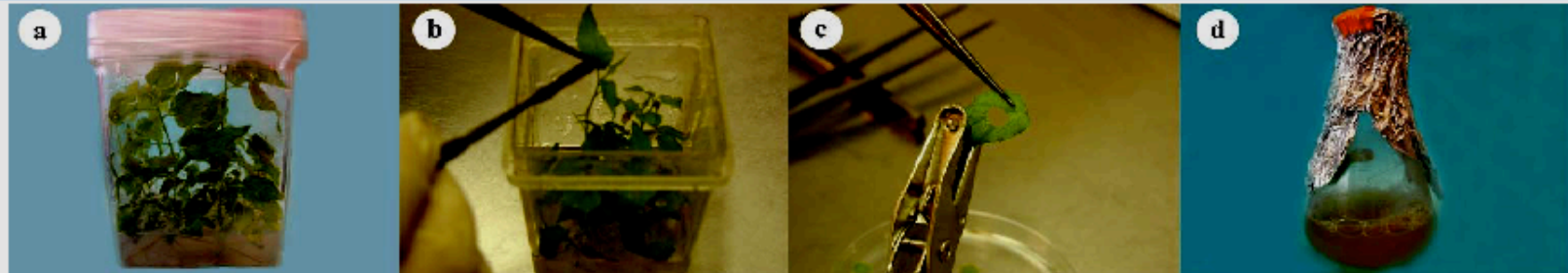


Il processo di trasformazione



Fasi della trasformazione

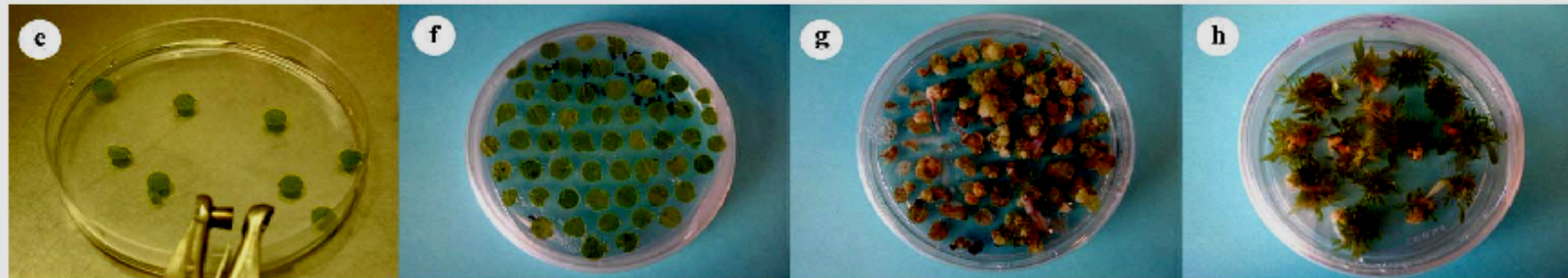


a Non-transgenic plant in a Magenta box. Source of explants for co-cultivation.

b Excising a leaf from an *in vitro*-grown plant. Done in a laminar flow hood.

c Punching leaf discs. Done with sterile implements in a laminar flow hood.

d Overnight (stationary-phase) culture of *Agrobacterium tumefaciens*.



e Leaf discs floated on *Agrobacterium tumefaciens* suspension.

f Leaf discs aligned on callus-induction media (CIM) after co-cultivation. Photo taken 2 days after plating. Note bacterial growth (white) on margins of discs.

g Callus production on leaf disc explants. Green sectors represent cells that survived exposure to the selection agent.

h Shoots induced on leaf disc explants. Done in the presence of a selection agent.



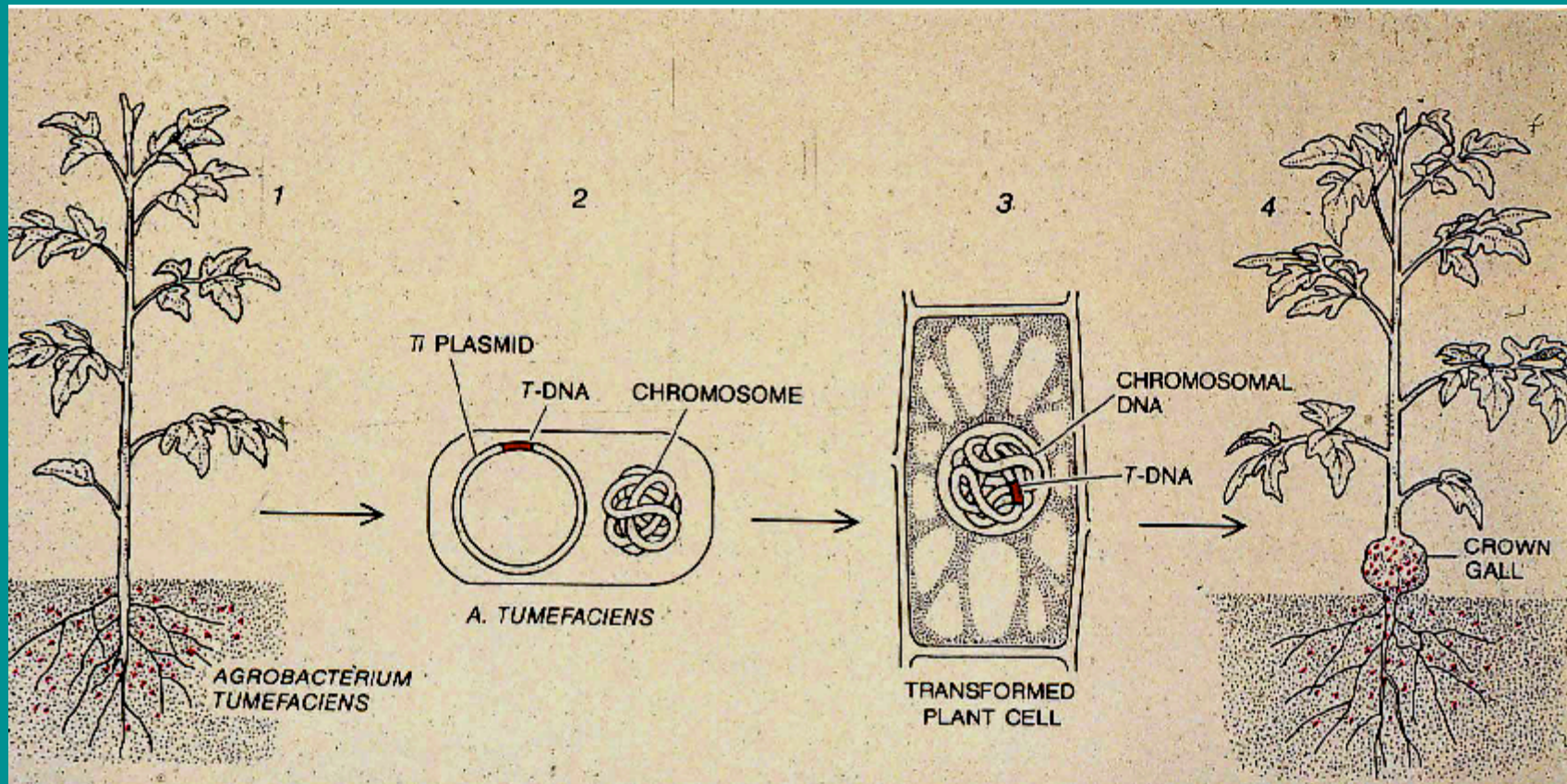
i Excised shoots inserted into root-induction media (RIM).

j Roots beginning to form on excised shoots. Done in the presence of a selection agent.

k Extensive root production on excised shoots.

l Fully regenerated transgenic plants growing in a Magenta box.

“Ingegneria genetica” naturale



Esempi di applicazione

- Resistenza agli erbicidi
- Resistenza ai patogeni (funghi, batteri, virus)
- Resistenza ai parassiti (insetti, nematodi)
- Resistenza agli stress abiotici (siccità, salinità del terreno)
- Miglioramento delle rese di produzione
- Fissazione biologica dell'azoto
- Decontaminazione dei suoli

Alcuni esempi per categoria

Classificazione

- Tolleranza a stress abiotico
- Tolleranza alla salinità
- Proteine di riserva del seme
- Qualità nutrizionale (provitamina A)
- Maschiosterilità o fertilità
- Fotosintesi
- Nodulazione
- Resistenza a funghi
- Protezione da virus
- Resistenza agli insetti
- Resistenza agli erbicidi

Gene chimerico

gpat, sod, MtID

betA, p5cs, hval, codA, afp, imt1

Phaseolin, phytohemagglutinin, conglycinin, patatin, zein, glutenin

psy, crtI, lcy

Ribonucleasi (barnasi), Inibitore della ribonucleasi (barstar)

Proteine che legano la clorofilla a/b, ribulosio-1,5-bisfosfato carbossilasi

Lectina, legemoglobina

Chitinasi, *ribosome inactivating protein* (RIP)

Proteine del capsid, RNA satellite

Tossina Bt, inibitori delle proteinasi

aroA e EPSP (glifosate), *bar* (fosfinotricina), *bxn* (bromoxynil)

Resistenza agli insetti: la tossina Bt

- *Bacillus thuringiensis* (Bt) colonizza naturalmente i suoli
- Produce proteine sotto forma di cristalli (protossine) che risultano tossici per le larve di alcuni insetti (lepidotteri, coleotteri, ditteri) quando sono ingeriti
- E' stato usato per più di 50 anni come insetticida naturale. Da 20 anni si usano preparazioni di spore e cristalli
- E' caratterizzato da elevata specificità d'ospite
- Sono state isolati almeno 90 geni diversi codificanti le protossine
- Piante trasformate con il gene per la tossina risultano resistenti
- Primi esperimenti effettuati nel 1987 su tabacco e pomodoro. In seguito sono state ottenute molte altre specie da raccolto (cotone, mais, riso, patata)

Resistenza agli insetti: altre fonti di geni

Inibitori delle proteasi

- Sono proteine presenti in molte specie vegetali e fanno parte del sistema naturale della pianta di difesa dagli insetti. Agiscono inibendo le proteasi a serina (tripsina e chimotripsina) degli insetti. Sono degli “antimetaboliti”
- Sono inattivati dalla cottura quindi sono ritenuti sicuri per l'alimentazione umana
- Gene *CpTi*, inibitore della tripsina da *cowpea* (fava)
- Geni dello stesso tipo sono stati isolati da numerose specie vegetali

Incremento della produttività

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Update

TRENDS in Biotechnology Vol.21 No.5 May 2003

Transgenesis and yield: what are our targets?

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Plant metabolic engineering has been used to generate a wide range of transgenic lines in which specific metabolic steps have been targeted. Attempts to increase yield of some agronomically important crops using this approach have highlighted the inherent complexities of modulating plant metabolism. In light of the recent findings by Regierer *et al.* this article addresses the major challenges faced with respect to enhancing yield through transgenesis.

The first use of transgenesis in plants in the 1980s heralded the arrival of a powerful and exciting tool for the study of metabolic regulation and for crop improvement. Of particular interest from a commercial viewpoint was the potential for increasing yield thus making the alteration of carbon partitioning between sucrose, starch and amino acids an important target for manipulation.

During the past 15 years a high proportion of the transgenic lines that have been generated has highlighted the problems associated with effective manipulation of plant metabolism. Plant metabolism displays plasticity

assumptions as to which factors might be important for metabolite partitioning based on our 'classical' views of metabolic pathways. Some improvements in yield have been achieved but the success rate has been low compared with our initial expectations.

Identifying targets

Several approaches have been taken to increase yield using transgenic technology. These include manipulating metabolism in source tissues with the aim of increasing carbon supply to heterotrophic sink tissues [6]; increasing transport capacities between source and sink tissues to try to increase photoassimilate supply to the sink tissues [7] and manipulating metabolism in the sink tissue to increase the utilization of photoassimilates thereby increasing amounts of specific compounds [1]. These approaches used either ectopic overexpression of heterologous genes or antisense repression of genes. Molecular biology, trait mapping and targeted gene reduction using transgenesis coupled with metabolic control analysis have