

Joint Research Centre (JRC)

Standardization of cTnI: Aspects to be considered for a model which will (is expected to) work



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<http://www.jrc.ec.europa.eu/>

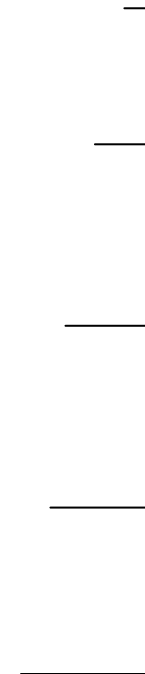
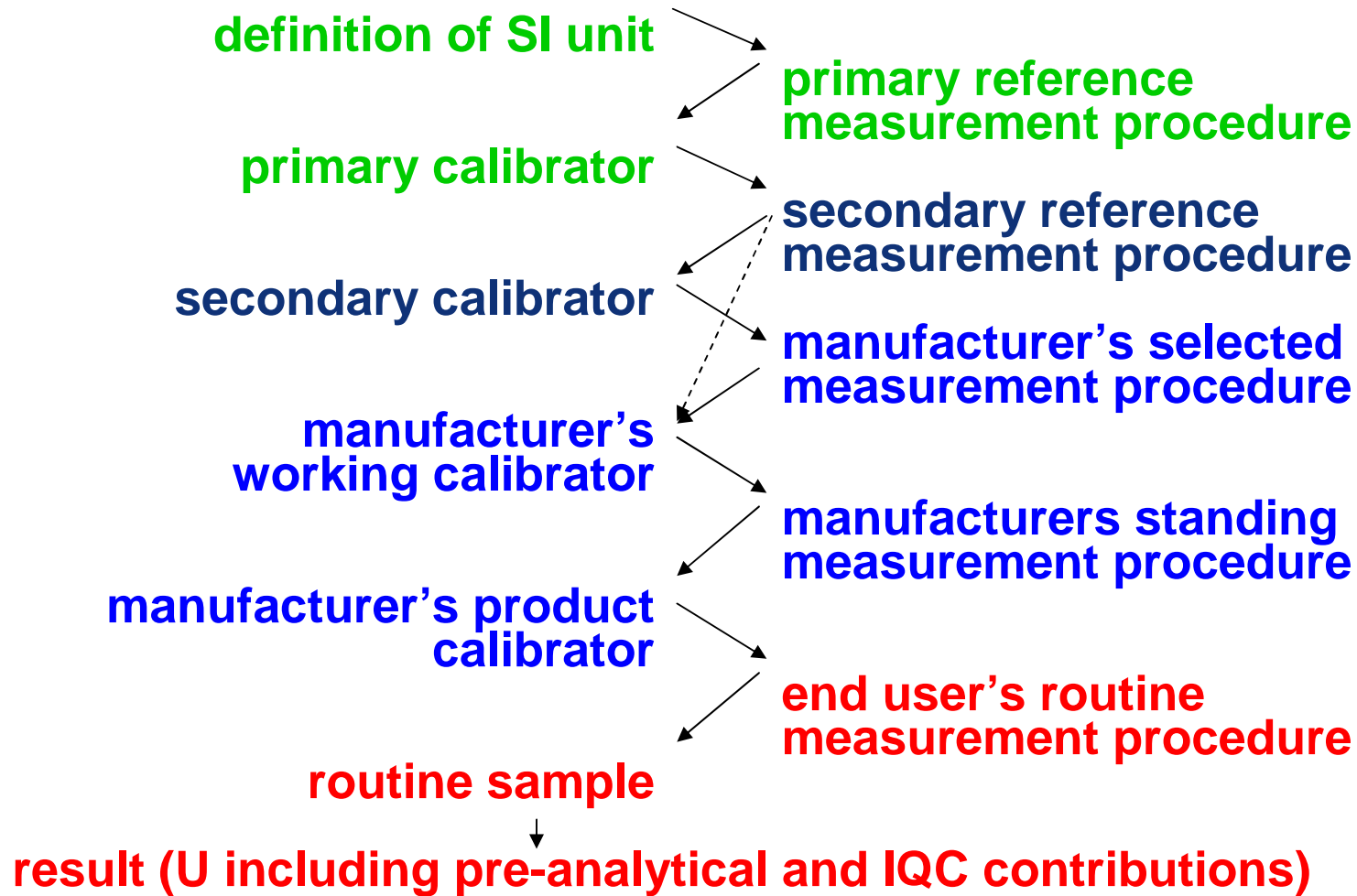
Definition of **common reference ranges** or **decision limits** for eventually critical diagnostic decisions

According to EQAS schemes for proteins **inter-assay variability tends to be relatively high**, in particular for TnI

Diagnostic power of particular biomarkers can eventually **not be exploited** to the full extend

Standardisation taken up in **legislation** (IVD-MD Directive 98/79/EC) and by ISO (ISO 17511 / ISO 18153) requiring ‘traceability to higher order reference methods and/or materials’

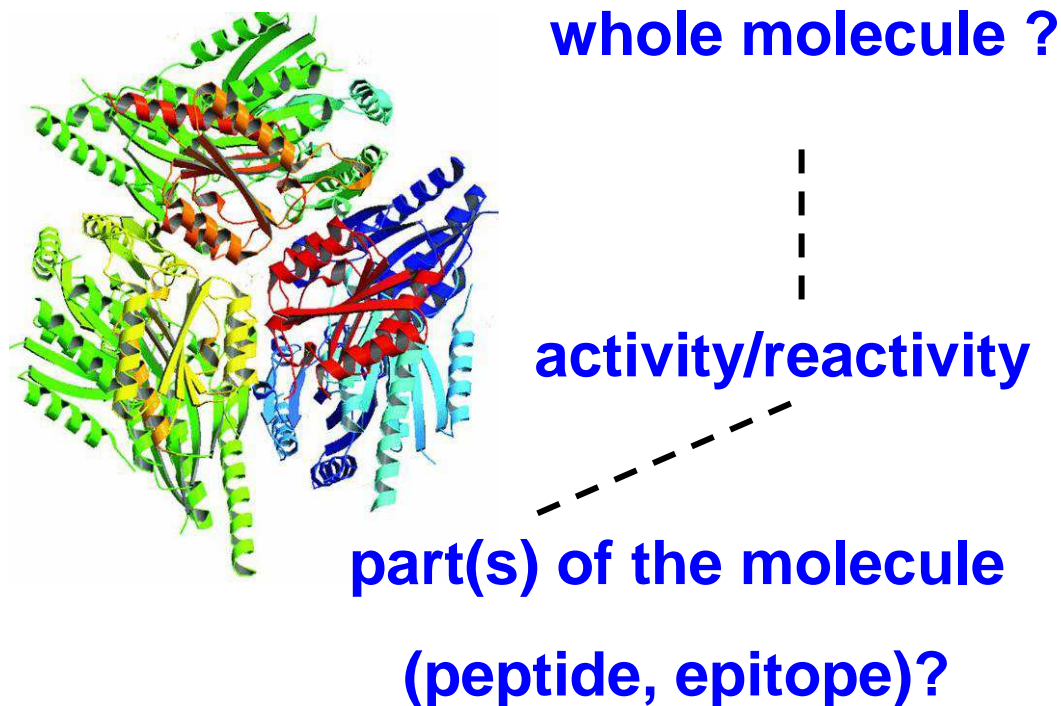
Uc



- **Metrological traceability to a stable reference, i.e. SI or a stable materialized standard**
- **Traceability via an unbroken chain of unbiased comparisons with known uncertainty**
- **Only then comparable measurement results can be achieved over space and long time periods**
- **Not a purpose on its own but means to achieve comparability of measurement results!**

To make sure that **traceability is not a purpose on its own** but leads to comparability of measurement results:

- **Definition of the measurand (VIM2) - VIM3 definition of measurand ('quantity intended to be measured')** ambiguous in this context
- **Calibrators (stable, homogenous) commutable** for routine methods and reference methods
- **Elimination of bias and control of all parameters which may cause bias**
- **Maintain the measurand (VIM2) or know the relationships through traceability chain (equal relationship in patient samples and calibrators required, otherwise traceability chain broken)**



**“straight forward”
for:
total amount of
defined molecules**

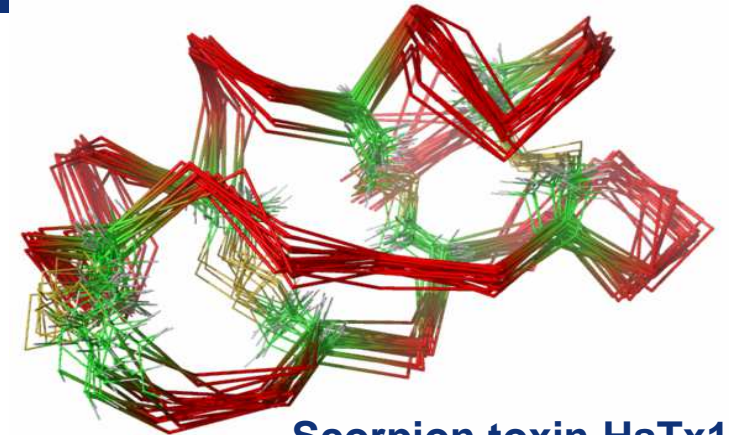
**Less evident:
amount of a protein
occurring in various
forms / fragments**

Problem: Defining the analytical target & unit

- The critical parameter is not only the “amount of substance”, **rather functionality** of the molecule or its parts in analytical systems that **matters (not identical to biological activity)**

Covalent:

- sequence (isoforms)
- splicing
- degradation (N-terminal, ...)
- chemical modification (oxidation, deamidation...)

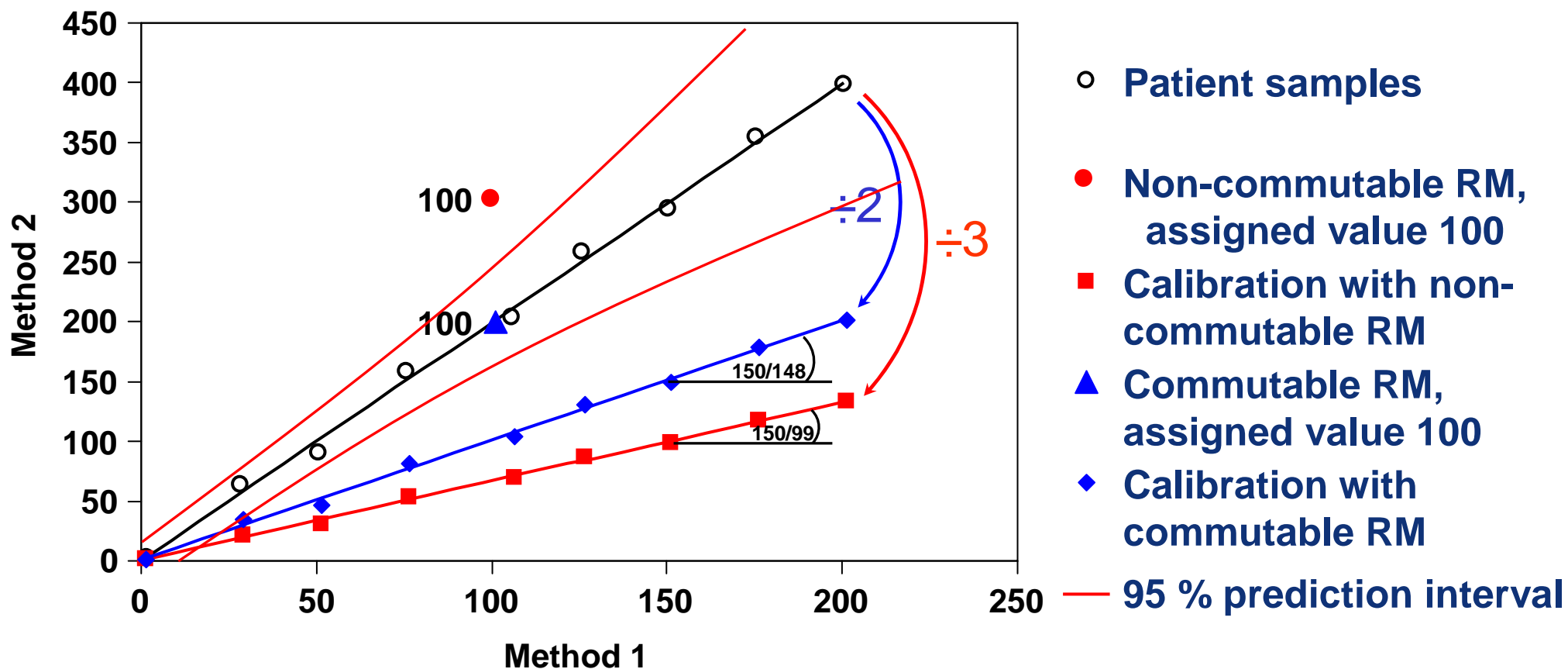


Scorpion toxin HsTx1
(NMR)

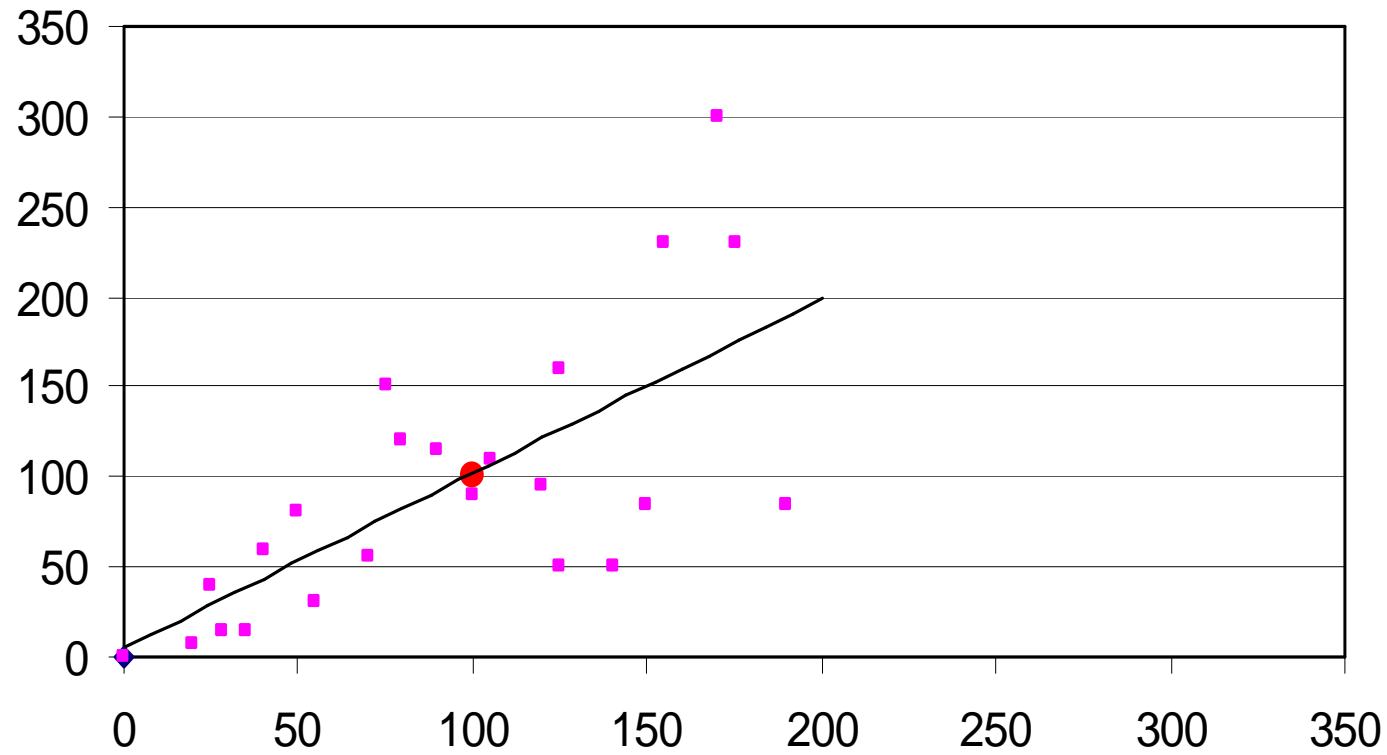
Non-covalent:

- oligomeric state
- ligand binding (metals, other proteins, co-factors, ...)
- degree of structuration (partial denaturation)
- different conformational states, unstructured proteins
- aggregation

- Proteins are typically quantified by measuring the amount of substance of a **certain part of all isoforms or of particular isoforms (epitopes for immunoassays, peptides for mass spectrometry methods)**
- A very **specific and strictly species-limited definition** of the measurand would mean that the **measurand** of the different methods would mostly be **different** without necessarily having different clinical significance
- **By definition results on different quantities cannot be compared**
- **Correlating measurands** can be harmonised and standardised

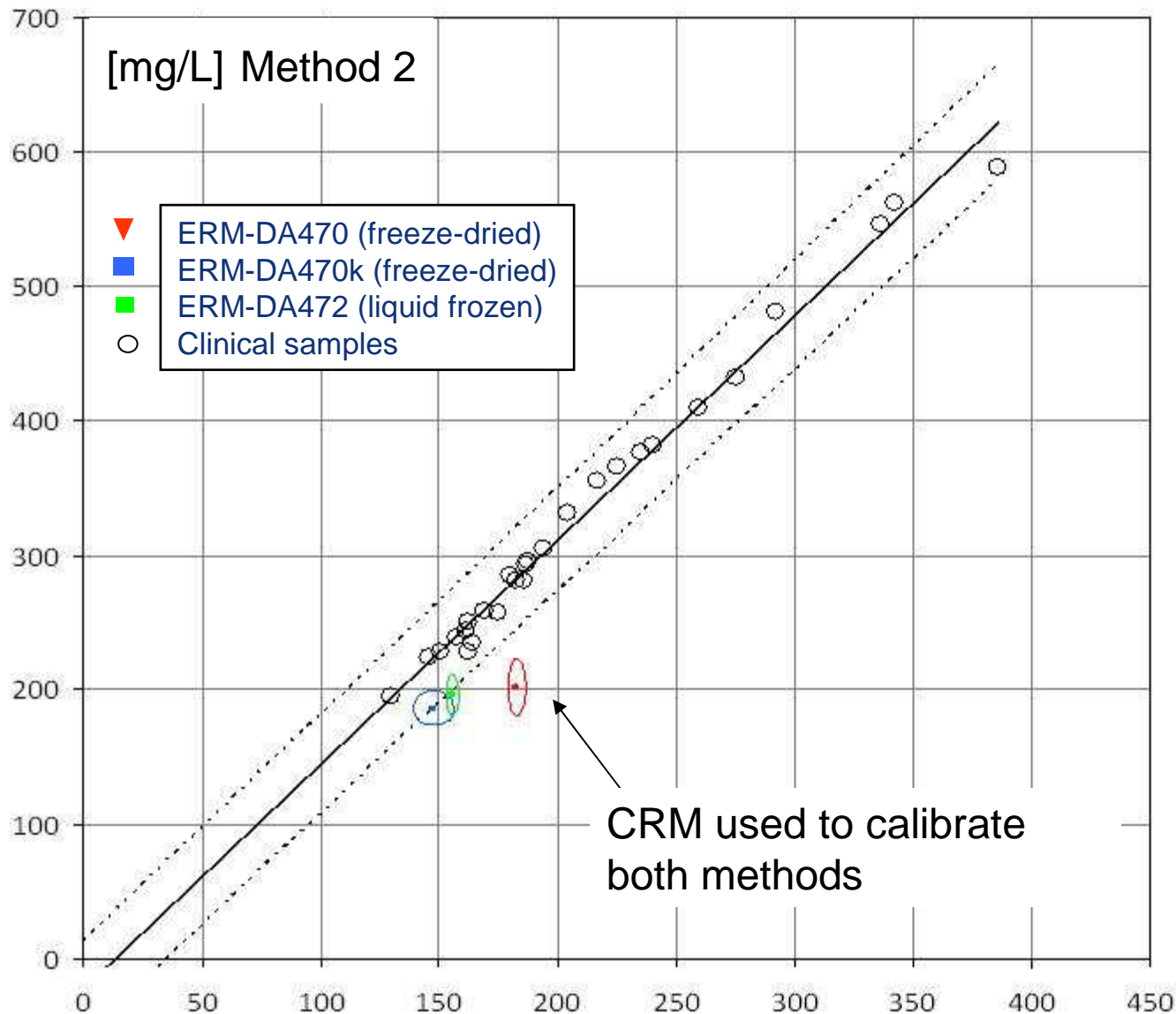


- Patient samples
- Non-commutable RM, assigned value 100
- Calibration with non-commutable RM
- ▲ Commutable RM, assigned value 100
- ◆ Calibration with commutable RM
- 95 % prediction interval



- **Lacking correlation reveals different method selectivity and non-constant relationship between different measurands of the methods applied**
- **Only averages of sample populations could be harmonized**
- **Pronounced differences between results for individual routine samples will persist after recalibration**
- **If data scatter is not acceptable the question has to be clarified which method gives the more clinically significant results and accordingly the measurand should be defined**

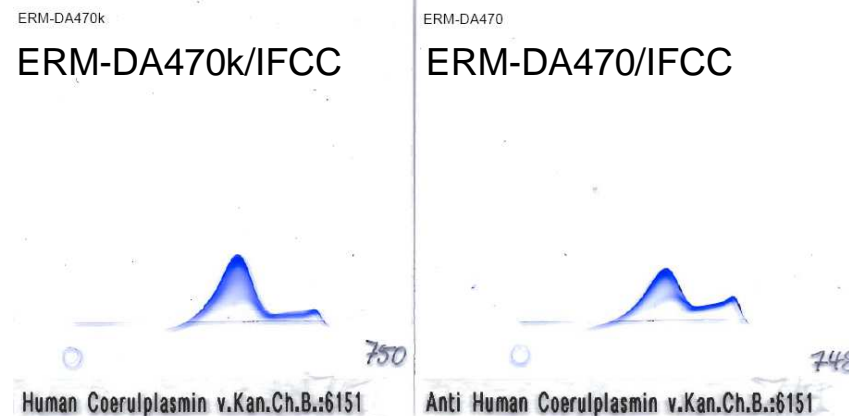
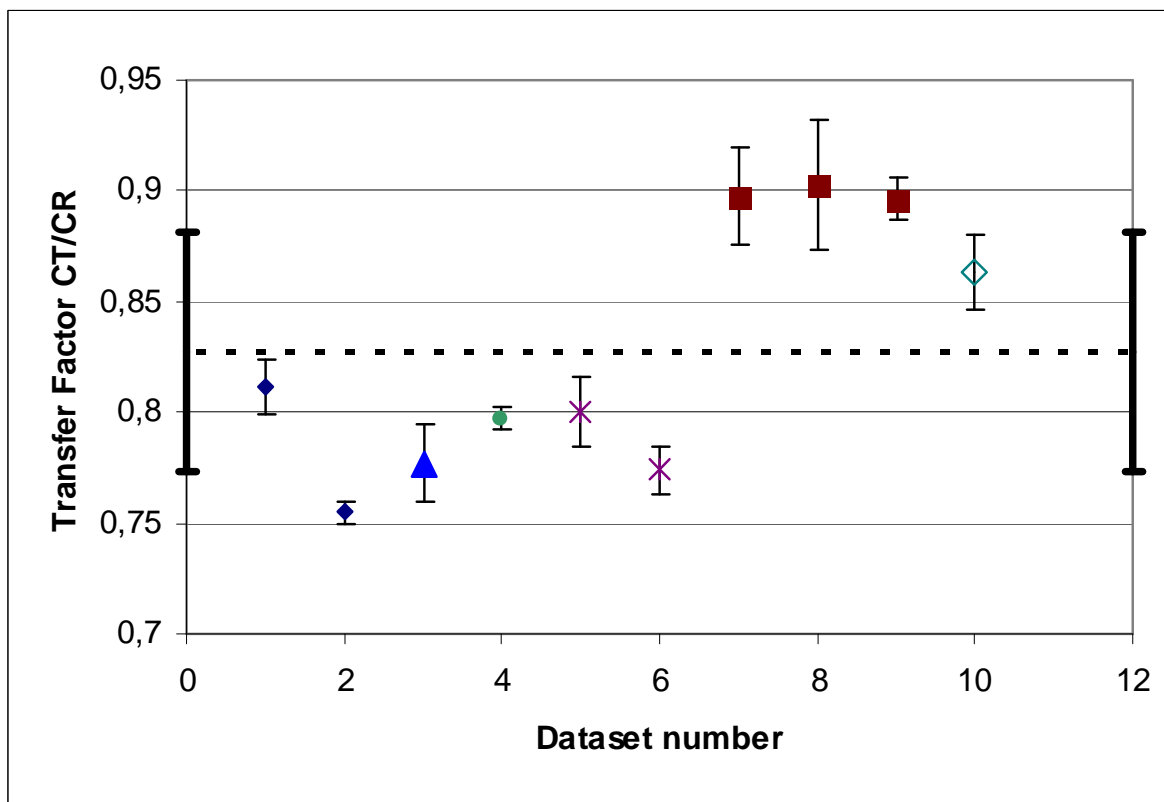
- **Reasons for lacking commutability**
 - **Matrix interactions**, differences between routine samples and reference material
 - Differences in method selectivity. Although routine methods may correlate well on typical routine samples the **presence of atypical isoforms or isoform patterns**, differently detected by routine methods in the reference material, may cause (eventually uncontrolled) bias

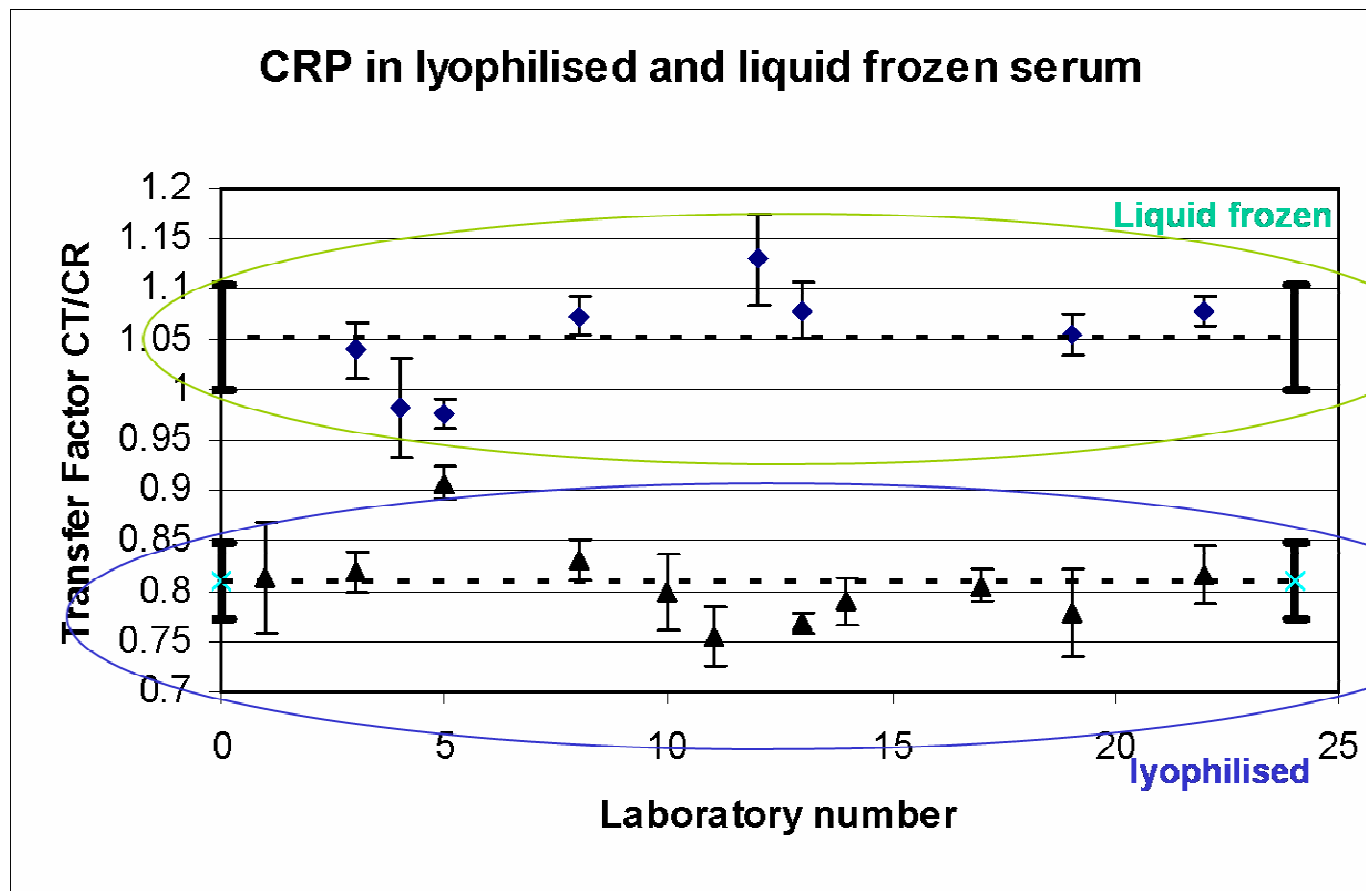


- Both methods were **using ERM-DA470** for calibration
- **Result** for ceruloplasmin in ERM-DA470 is reasonable with both method 1 and 2 (certified concentration 205 mg/L)
- **ERM-DA470 is not commutable** for this combination of methods



Discrepant results for clinical samples

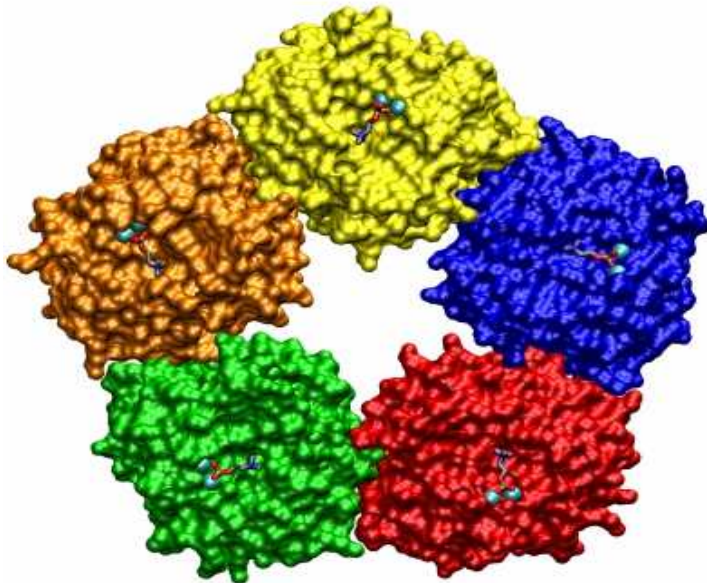




The issue: Lyophilisation results in a loss of about 20 % of measurable CRP compared to the non-lyophilised material and both formats appear commutable!

Problems observed for CRP:

- Only 80 % detected by routine assays after freeze-drying
- High between bottle heterogeneity in freeze-dried material
- Presence of different oligomeric forms in freeze-dried material

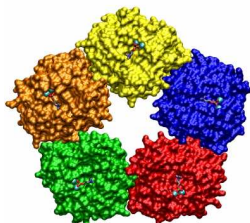


Pentameric protein
Monomer: 25 106 Da
Binds two Ca²⁺

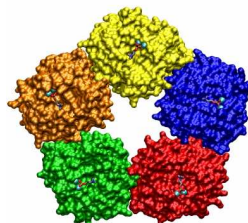
Physicochemical state verified by:

Gel filtration followed by SDS PAGE and Western blotting

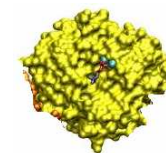
Semi-native gel electrophoresis followed by Western blotting



Pentamer in serum matrix



Pentamer in buffer



Monomer in buffer

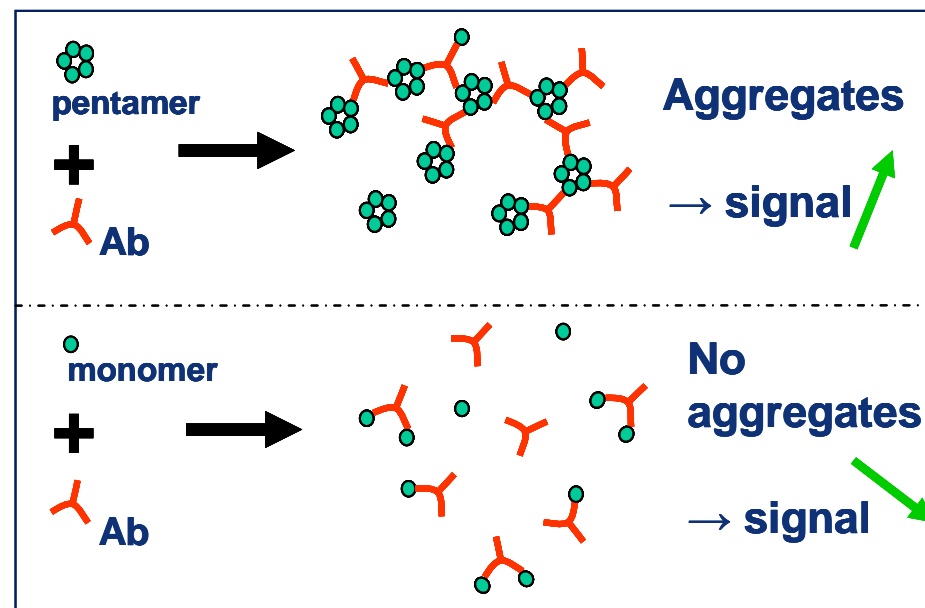
| Material | nominal conc | CRP mass concentration measured with different methods (n=3), relative to the nominal concentration | | | | | | | |
|--------------------|--------------|---|-------|-------|-------|-------|-------|-------|-------|
| | | IA1 % | IA2 % | IA3 % | IA4 % | IA5 % | IA6 % | IA7 % | IA8 % |
| ERM-DA472/IFCC | 100 | 101 | 89 | 107 | 100 | 89 | 98 | | 100 |
| Pentamer In buffer | 100 | 103 | 49 | 59 | 79 | 96 | 118 | | 100 |
| Monomer In buffer | 100 | 12 | 1 | <1 | <13 | <8 | 2 | 3 | 2 |

- mCRP gives < 10 % response in homogeneous immunoassays compared to nCRP

The loss in measured CRP found for ERM-DA470k/IFCC is due to the partial dissociation of CRP into monomers which are not recognised
The presence of 2 mM calcium can prevent the dissociation
Liquid frozen ERM-DA472/IFCC does not contain detectable amounts of mCRP

M. Rzychon, I. Zegers, H. Schimmel Analysis of the physico-chemical state of CRP in different preparations including two reference materials *Clinical Chemistry* 2010;56:1475-82

**Impact of the oligomeric form on the homogeneous immunoassay response?
Impact depends on immunoassay format!**



Method or instrument:

Beckman Immage

Dako

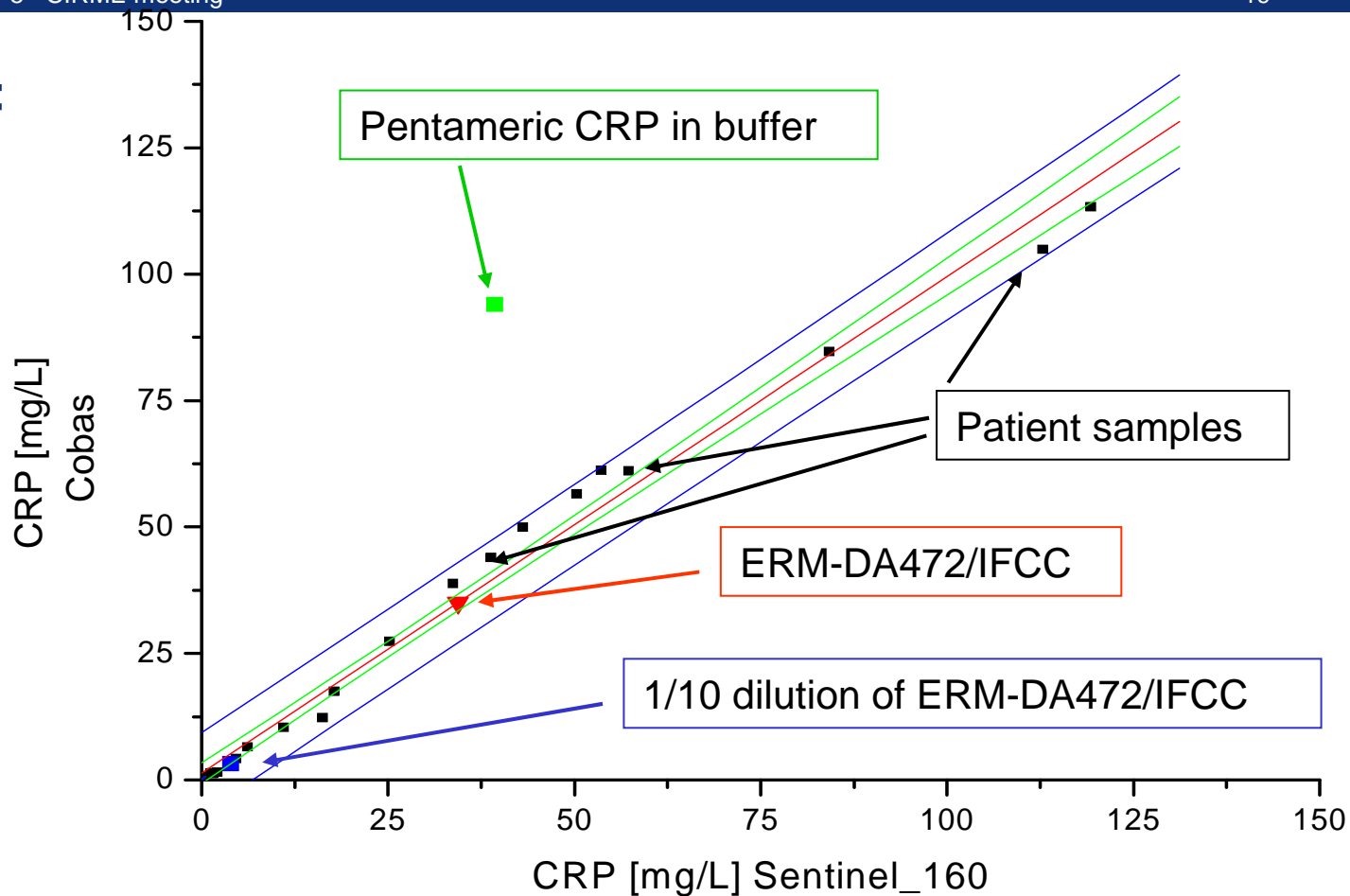
Sentinel

Abbott Architect

Cobas

Integra

BN ProSpec

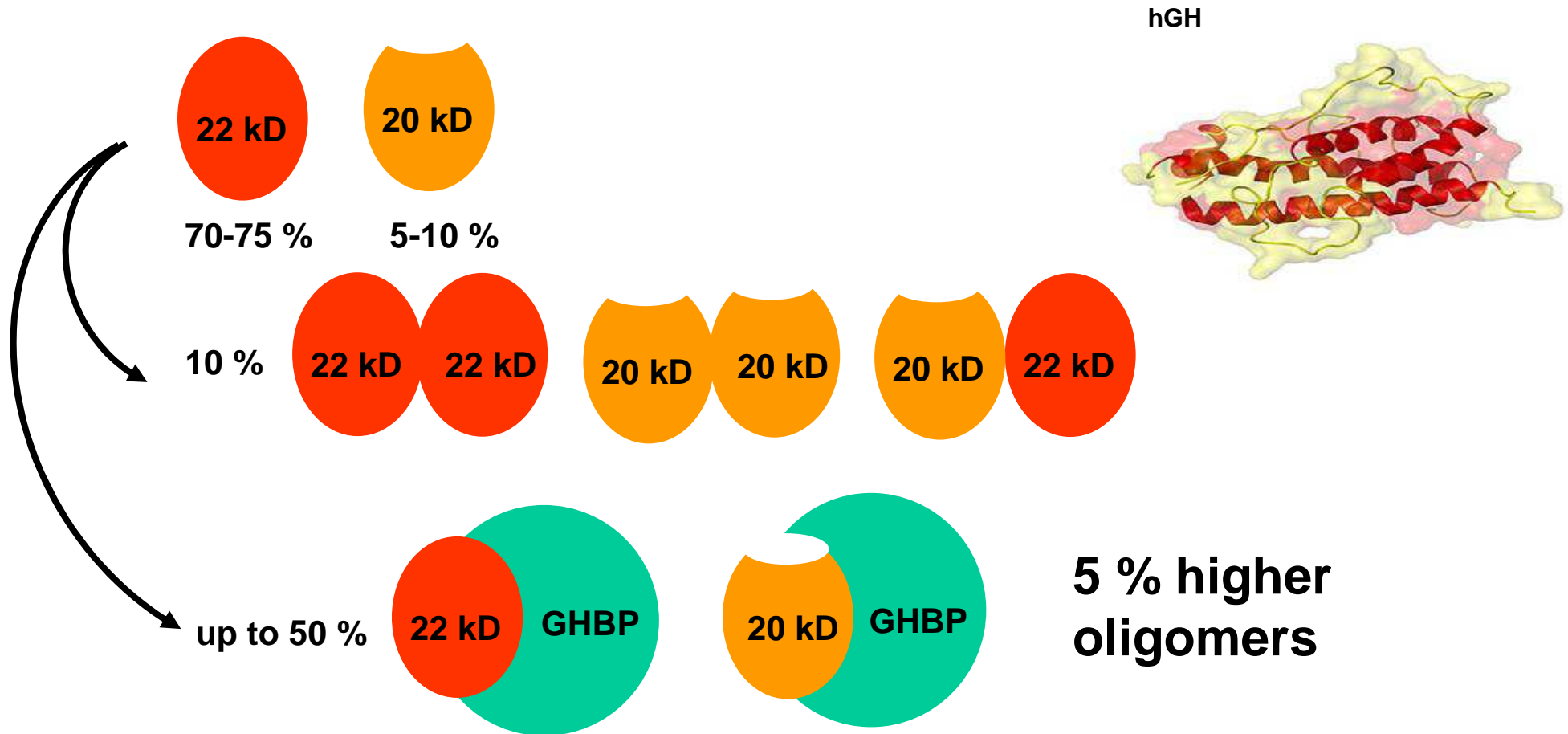


ERM-DA472/IFCC is commutable

A 1/10 dilution of ERM-DA472/IFCC is commutable

CRP without matrix is not commutable















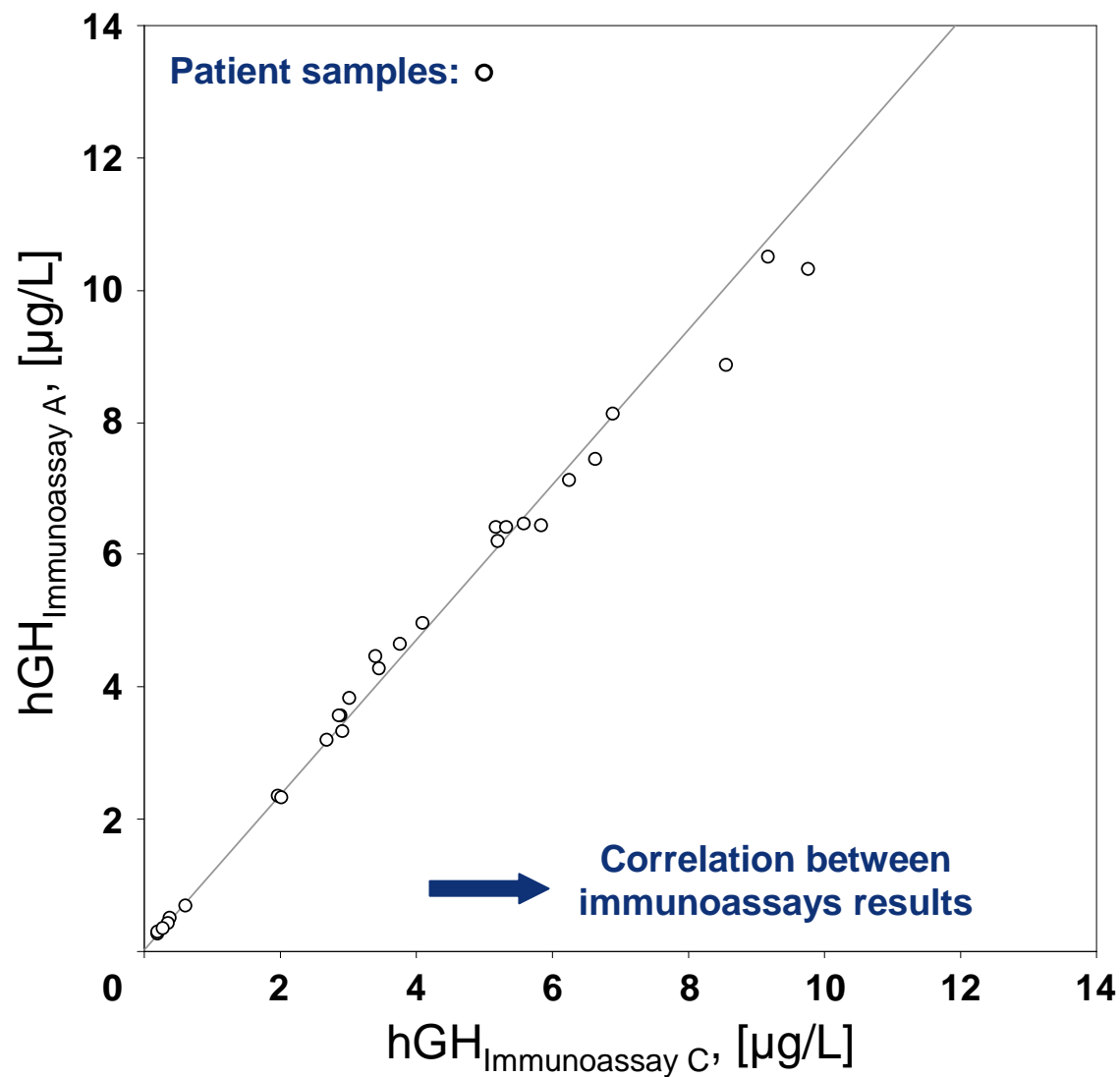
Methods evaluated: Main routine immunoassays

Immunoassay C versus A

| Manufacturer | Calibration | Symbol |
|--------------|-------------|--------|
| Siemens® | WHO 98/574 | A |
| Roche® | WHO 98/574 | B |
| Dia Sorin® | WHO 98/574 | C |
| Mediagnost® | WHO 98/574 | D |
| IDS® | WHO 98/574 | E |

| Immunoassays | A | B | C | D | E |
|--------------|---|--|---|---|---|
| A | |  |  |  |  |
| B | | |  |  |  |
| C | | | |  |  |
| D | | | | |  |
| E | | | | | |

➔ 10 comparisons



Methods evaluated: Main routine immunoassays

➔ 10 comparisons

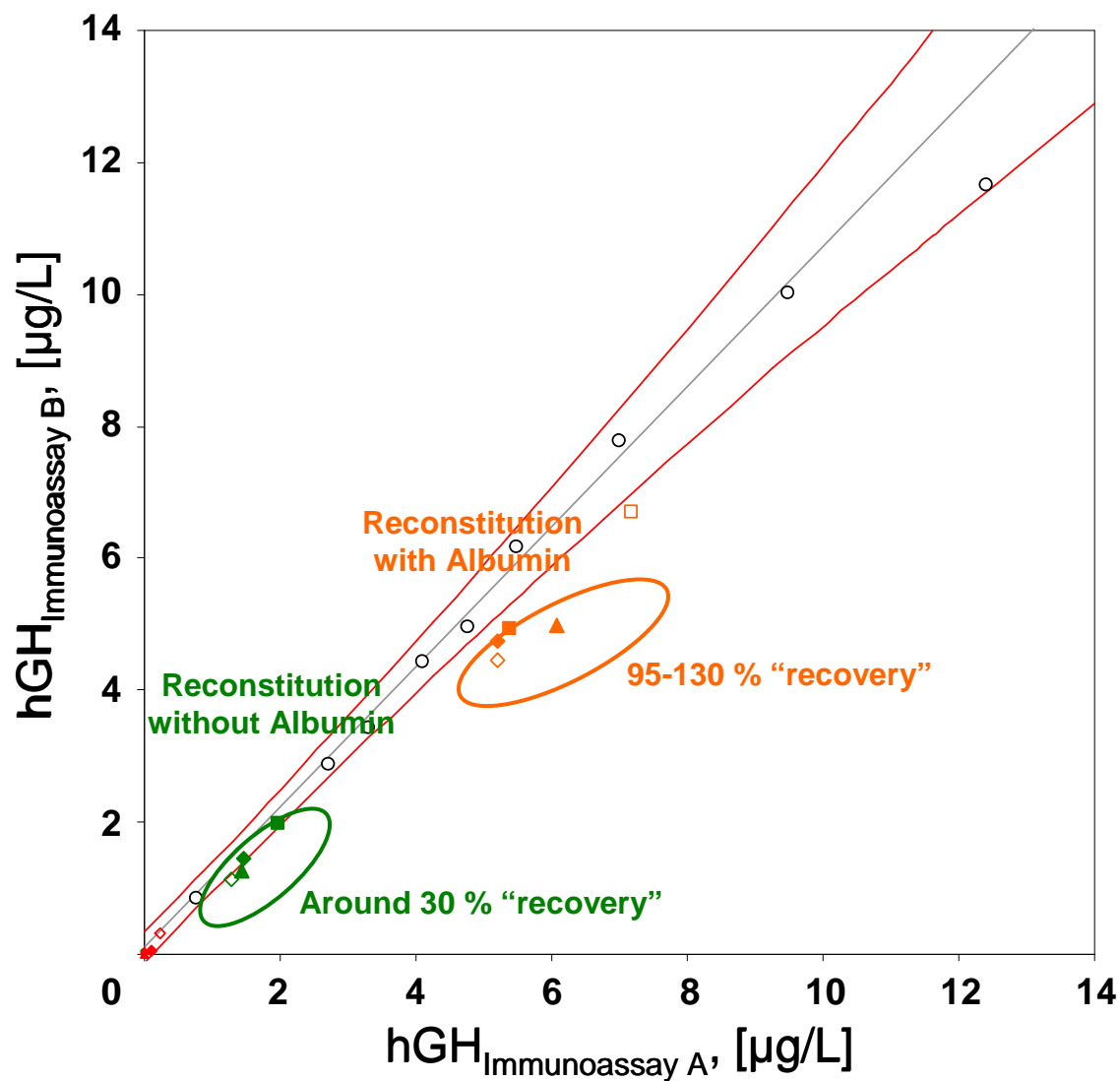
Patient samples: ○

Reconstitution: ■ Phys. buffer
 ■ Phys. buffer - Albumin

Blank controls: ■

Spiking: ○ Phys. buffer
 □ Phys. buffer - Albumin
 ■ hGH depleted serum
 ◆ Charcoal stripped serum
 ◇ SRM 971 male
 ▲ Sheep serum

Immunoassay A versus B



- **Measurands have to be strictly defined or need to correlate for the routine and reference methods in order to allow harmonization / standardisation**
- **Methods determining the amount of substance of a protein only (such as mass spectrometry on peptides or the protein backbone) may not be sufficient to control the traceability chain well enough (see CER, CRP, hGH)**
 - A well understood immunoassay based reference method is a valid option for standardizing methods with equal specificity and/or correlating measurands

- **Control (and quantification) of all relevant influence parameters with the related uncertainties (e.g. matrix properties and interaction, structure, aggregation, oligomerisation, isoform profile, metal binding) is a prerequisite for being able to reproducibly produce reference materials / calibrators and to use them for calibration in a way that comparable measurement results can be achieved on long term.**
 - Highly fluctuating isoform patterns in patient samples require equal response of the routine methods and in particular of the reference method to allow for a maximum degree of harmonisation / standardisation

- **Non-commutability** of a reference material means that using it for calibration **will introduce a bias** for at least one of the methods, whereas **commutability** means that it can be used for **achieving comparable results** but it does **not guarantee the absence of bias** (see **CRP and hGH**). **Commutability is absolutely required!**
- Careful evaluation of the **impact of processing** steps on the properties of quality control materials / PT samples / reference material is crucial
- If necessary **uniform calibration protocols** using commutable calibrators and **commutable dilutions** thereof need to be developed (e.g. for the calibration of **high sensitivity assays**)

- **A higher order reference material or reference measurement procedure cannot stand alone, they are integral parts of reference measurement systems**
- **Application of quality control / reference materials outside their validated intended use requires verification of their usability for the extended purpose**
- **Standardisation of protein biomarkers is challenging, however, often possible with reasonable efforts and within acceptable timelines**
- **Correlation is key!**

Collaborative research project on the traceability of values for complex biomolecules:

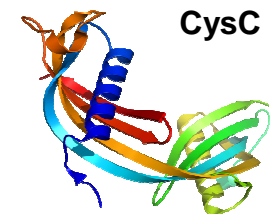
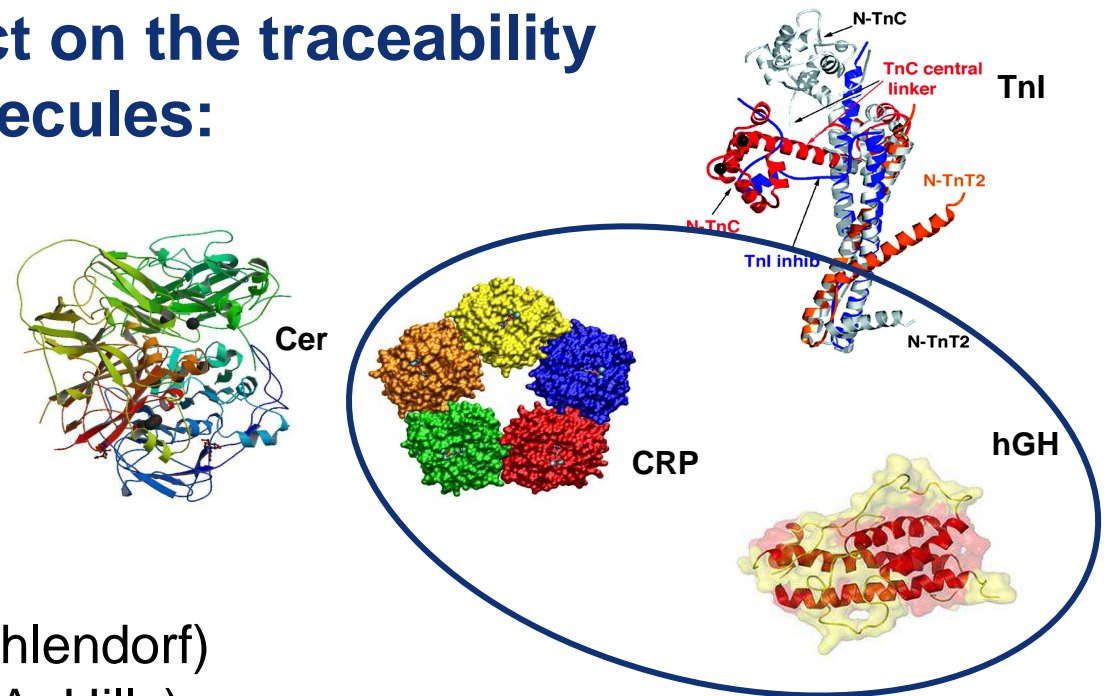
‘amount of substance’



Functional activity

Partners and collaborators:

- PTB (A. Henrion, C. Arsene, R. Ohlendorf)
- NPL (M. Ryadnov, P. Rakowska, A. Hills)
- LGC (G. O’Connor, M. Quaglia, H. Parkes, C. Pritchard, S. Biesenbruch)
- IRMM (I. Zegers, H. Schimmel, A. Munoz-Pineiro, M. Ryzchon, G. Auclair, K. Hanisch, S. Boulo)
- IFCC-WG hGH: C. Sturgeon, M. Bidlingmaier



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